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Acute stressor-selective effect on total plasma homocysteine concentration in rats

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Abstract

Stress produces several physiological and behavioral alterations that increase cardiovascular morbidity and mortality. Nonetheless, there is a dearth of studies that have evaluated the effects of stress on total plasma homocysteine, an important amino acid associated with cardiovascular disease. We used four distinct acute stressors in rats, i.e., swimming, restrain, novelty and cold exposure, in order to examine whether any acute effect on total plasma homocysteine concentrations would occur. Plasma corticosterone and adrenocorticotropic hormone concentrations were also measured to demonstrate the ability of the chosen manipulations to activate the hypothalamic–pituitary–adrenal (HPA) axis. Three of the four stressors activated the HPA axis and only restrain affected total plasma homocysteine concentrations (+37%, P=.006) compared with the control group. The complexity of the physiological responses to stress, the peculiarities of stress responses and the intricate regulatory systems involved in homocysteine metabolism must be taken into account in order to clarify the increasing effect of restrain (mainly a psychological stressor) on total plasma homocysteine concentrations in rats and to evaluate its meaning in human pathology.

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1. Introduction

Epidemiological studies show that stress has an important impact on human health. Of particular interest are the stress-induced physiological and behavioral alterations that could influence, either directly or indirectly, the function of the cardiovascular system, and thus increase cardiovascular morbidity and mortality (Møller et al., 1996; Fauvel, 2002; Ramey, 2003). Both human and animal studies (Blumenthal et al., 1995; Giorgiades et al., 2000, Yudkin et al., 2000, Muller et al., 2001; Black and Garbutt, 2002) have provided findings on the mechanisms by which stress interferes with immune, neuroendocrine and metabolic changes that may increase cardiovascular risk, even in the absence of other major risk factors (Manuck et al., 1995).

Homocysteine is a sulfur-containing amino acid intermediate in methionine-cysteine metabolism (Finkelstein and

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Martin, 2000), as it represents a branching point at which it can be remethylated to methionine or converted to cysteine. Plasma homocysteine concentrations are dependent on complex metabolic regulation, including hormone, redox and possibly immune mediators, as well as on availability of nutrients (mainly folate, B6, and B12 vitamins) that serve as cofactors for metabolic reactions (Tobeña et al., 1996; House et al., 1999; Stead et al., 2000; Graham and O'Callaghan, 2002; Vitvitsky et al., 2003). Some of these regulatory systems are activated by stress and could mediate the changes in homocysteine purported to occur in such situations.

Increase of total homocysteine plasma concentrations (the sum of protein-bound, free and disulfide fractions) is now recognized as an independent risk factor for cardiovascular disease (Nygård et al., 1999). Although the strongest associations were demonstrated in case–control and cross-sectional studies, a recent meta-analysis of prospective studies (Hackam and Anand, 2003) also showed that in specific conditions, such as diabetes, chronic renal disease and systemic lupus erythematosus, hyperhomocysteinemia appears to be a predictive factor of mortality (Guilland et al., 2003).

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Very few studies have addressed the effects of stressful manipulations on total plasma homocysteine concentrations. To the best of our knowledge, there is only one study that reports that one session of psychological stress increases plasma homocysteine concentrations (Stoney, 1999), while another did not find any such changes in women (Farag et al., 2003).

In the present study four distinct stressful manipulations were employed. Rats were restrained (considered mainly as a psychological stressor), exposed to swimming (a stressor that interferes with cardiovascular and metabolic homeostasis), cold (physical/metabolic stressor) or to novelty (psychosocial stressor) in order to verify whether stress, in general or in particular, would have any acute effect on total plasma homocysteine concentrations. We also measured corticosterone (CORT) and adrenocorticotropic hormone (ACTH) concentrations as control measures to index the extent to which the animals were stressed.

2. Methods

2.1. Animals

Fifty male, 3-month-old Wistar rats from the animal facility of the Department of Psychobiology were used. They were distributed into five groups corresponding to the different stress manipulations (restrain, swimming, cold, novelty and control). Each stress session, except for swimming, lasted 1 h, beginning approximately at 2:00 p.m. Swimming sessions lasted 40 min because at this point, animals began to drown. Therefore, we established this as the maximum swimming time in order not to make the animals hopeless (an animal model of depression). Animals were sacrificed immediately after the end of each session. All procedures used in the present study complied with the National Institutes of Health *Guide for the Care and Use of Laboratory Animals*.

2.2. Experimental procedures

2.2.1. Control group

Animals were maintained in their original home cages in groups of five rats.

2.2.2. Restrain

Ten animals were individually placed inside plastic cylinders (21 cm in length \times 6 cm in diameter). Both ends of the cylinders were closed with ventilation sliding doors.

2.2.3. Swimming

Animals were placed in a container filled with water at a height of 50 cm. Tap water was not heated, being approximately 20 $^{\circ}$ C.

2.2.4. Cold

Ten animals were individually placed in a cold chamber at 4 $^\circ \rm C$ inside a wire-mesh cage.

2.2.5. Novelty

Animals were individually placed in a cage filled with used sawdust taken from another cage.

Immediately after the end of each stress session, animals were brought into an adjacent room and decapitated. Trunk blood was collected in precooled plastic vials, containing 0.2 ml of an EDTA solution (60 mg/ml) and centrifuged at 900 × g for 20 min at 4 °C. Plasma was extracted, transferred to plastic tubes and stored at -80 °C until biochemical assays were performed.

2.3. Assay procedures

2.3.1. Homocysteine determination

Total plasma homocysteine values were determined by high-performance liquid chromatography (HPLC) with fluorimetric detection and isocratic elution (Pfeiffer et al., 1999). This methodology involves three steps, namely, reduction of thiol groups using tris(carboxyethyl)phosphine, protein precipitation and derivatization with 7-fluorobenzene-2-oxy-1,3-diazolic-4-ammonium sulfate-SBD-F. The HPLC system used was a Shimadzu apparatus with a SIL-10Dvp automatic sample injector and an RF-10AXL fluorescence detector. Chromatographic separation was performed using a C18 model Shim-pack CLC-ODS column (4.6 × 150 mm, with 5.0-mm microparticles). The fluorescence of the separated compounds was detected with a detector adjusted for excitation at 385 nm and emission at 515 nm. Total homocysteine content was calculated with a calibration curve using known homocysteine concentrations and cystamine as the internal standard. The intra-assay coefficients of variation (CVs) for homocysteine ranged from 1.1% to 1.8% and the interassay CV was 5.6%.

2.3.2. ACTH and corticosterone determination

The ACTH assay was performed by a sequential chemiluminescence immunometric method (DPC Immulite ACTH, Los Angeles, CA), using a monoclonal murine antibody specific for ACTH. The sensitivity of the assay is 9 pg/ml, and intra- and interassay variations are 9.6% and 9.4%, respectively. Corticosterone concentrations were assayed by a double antibody RIA method, specific for rats and mice, using a commercial kit (ICN Biomedical, Costa Mesa, CA). The sensitivity of the assay is 0.25 μ g/dl, and intra- and interassay variations are 10.3% and 7.1%, respectively.

2.4. Statistical analysis

The results of each variable were compared using a oneway ANOVA followed, when ANOVA detected a significant difference among groups, by the Newman-Keuls test for pairwise comparisons. The level of significance was established at $P \leq .05$.

3. Results

Plasma ACTH concentrations (Fig. 1, upper panel): ANOVA revealed a statistically significant difference among groups [F(4,39)=27.2; P<.00001], the highest ACTH concentrations having been observed in animals subjected to swimming (P<.0002).

Plasma corticosterone concentrations (Fig. 1, middle panel): A significant difference among groups was revealed [F(4,41)=32.86; P<.00001]. The highest concentrations were found in animals exposed to swimming (P<.0002), followed by those submitted to restrain and cold stress,

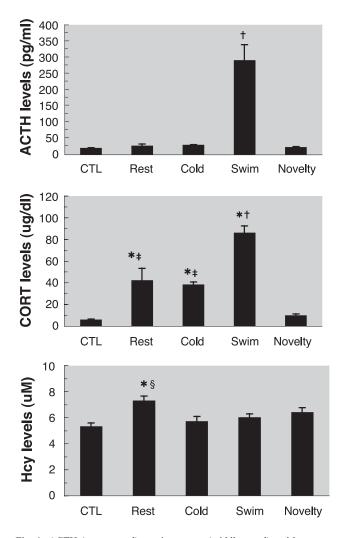


Fig. 1. ACTH (upper panel), corticosterone (middle panel) and homocysteine (Hcy; lower panel) plasma concentrations of animals submitted to different types of acute stress. The stressors used were restrain (rest), cold, swimming (swim) or novelty. Values are presented as mean \pm S.E.M. of 8– 10 animals/group. * Different from CTL group; †Different from all groups. ‡Different from novelty group. §Different from control (CTL) group.

which were equally higher than novelty-exposed and control animals (P < .001).

Plasma homocysteine concentrations (Fig. 1, lower panel): Total homocysteine plasma concentrations were significantly altered by stress [F(4,43)=3.70, P=.01], with the highest concentrations found in restrained rats (7.31 µM), as compared to cold-exposed (5.73 µM, P=.03) and control (5.30 µM, P=.006) groups. Swimming and novelty-exposed groups presented intermediate values, which were not statistically different from the remaining groups.

4. Discussion

As supported by increased activity of the hypothalamic– pituitary–thyroid (HPA) axis, all stress manipulations were effective except novelty. Homocysteine changes occurred in a stressor-specific manner, in that restrain, a model of psychological stress in rats, significantly increased the amino acid concentration (+37%, P=.006). There were modest increases in the swimming (+13%) and novelty (+20%) groups that did not reach statistical significance.

The homocysteine increase in the restrain group, a stress mainly considered of psychological nature represents an intriguing finding. Similar results were reported by Stoney (1999), who demonstrated a significant elevation (+7%) of plasma homocysteine in women in response to mental arithmetic and verbal tasks (10-min baseline period and 10-min stress period). In contrast, another recent study (3min preparation and 3-min stress period), found no effect of a verbal task on plasma homocysteine concentrations (Farag et al., 2003), possibly due to the shorter duration of the stress session in comparison to Stoney's work (1999) (3 vs. 10 min baseline and stress periods). Intrinsic psychological aspects of cardiovascular diseases, e.g., hostility and anger, have also been associated with increased plasma homocysteine concentrations in humans (Stoney and Engebretson, 2000). In the two studies that showed increased homocysteine concentrations, the results were attributed to elevated sympathetic activity of the individuals. Unfortunately our study was not designed to measure such parameters.

In a previous work from our laboratory, we reported that rats sleep deprived for 96 h had lower total plasma homocysteine concentration compared to home cage controls and that this concentration did not increase even when they were allowed to sleep for 24 and 48 h. Differently, rats sleep deprived for 24 h did not present altered homocysteine concentrations compared to controls (De Oliveira et al., 2002).

At first we attributed the differences between sleep deprivation and restrain stress to the duration of the stressor and hypothesized that acute and chronic stressors could have opposite effects on homocysteine concentrations. Although we originally supposed that decreased plasma homocysteine concentrations in sleep-deprived rats could be associated with the hypothyroid state induced by sleep deprivation, we now believe that these differences may represent the degree to which each of the possible regulatory systems involved in homocysteine metabolism are elicited by a specific stressor, and that duration is related to the latency of the physiological response or sequential activation of these regulatory factors.

Our study indicates that the nature of the stress manipulation is also important, as it may selectively activate the main regulatory systems involved in response to stress (Pacák and Palkovits, 2001). On one hand, stress activates the sympathetic nervous system (SNS) and the HPA axis. Thus, during stressful situations, norepinephrine (from the SNS) and glucocorticoids (from the HPA axis) are released. Stress is also generally associated with increased growth hormone, prolactin, ACTH, norepinephrine and epinephrine concentrations (Lenox et al., 1980; Meyerhoff et al., 1990; Pacak et al., 1998; Pacak and Palkovits, 2001). On the other hand, the effects of stress on the thyroid are variable. Although a review by Pacak and Palkovits (2001) states that restrain inhibits the hypothalamic-pituitary-thyroid axis, Armario et al. (1986) demonstrated increased thyroid stimulating hormone concentrations in rats after 15-min sessions of four different psychological stressors in rats. However, restrain stress was not evaluated.

It is noteworthy that the abovementioned stress-induced hormone changes may interfere with homocysteine metabolism, a fact that may explain the different stress-induced responses. There are some studies in the literature that showed endocrine regulation of homocysteine metabolism. For instance, glucagon has been shown to reduce plasma homocysteine by decreasing homocysteine export from cells and by increasing the activity of cystathionine beta-synthase (CBS) and cystathionine gamma-lyase (CGL) enzymes in the transulfuration pathway (House et al., 1999; Jacobs et al., 2001). Insulin also seems to increase plasma homocysteine concentrations by decreasing CBS and CGL and the remethylation methylenetetrahydrofolate reductase enzyme activities (House et al., 1999; Dicker-Brown et al., 2001). In addition, hypothyroid rats were also shown to present reduced plasma homocysteine concentrations (Jacobs et al., 2000) and it has been proposed that glucocorticoids alter plasma homocysteine concentrations by stimulating CBS expression (Ratnam et al., 2002). Gender-related plasma homocysteine differences also point to an influence of sexual hormones on homocysteine metabolism regulation (Dierkes et al., 2001), as men have increased plasma homocysteine concentrations compared to women, although the mechanisms are still unknown.

Much attention has been paid to the endocrine regulation of homocysteine metabolism, but other physiologic mediators are potentially involved in the changes of plasma homocysteine concentrations induced by specific stressors, such as the case of inflammatory mediators (Tobeña et al., 1996; Black and Garbutt, 2002), and the vasoactive peptide endothelin-1 (Palma et al., 2002; Er et al., 2002). In fact, the extent to which each one of these regulatory systems are activated by a specific stressor will determine the final observed change in homocysteine concentrations, which can be increased, decreased or unaltered.

Particular stressors are associated with hyperhomocysteinemia, which may contribute to the deleterious effect of modern life stress. The complexity of the physiological responses to stress, the peculiarities of stress-selective responses, and the intricate regulatory systems involved in homocysteine metabolism should be better studied so that conclusions concerning the restrain-selective increase in total plasma homocysteine concentrations in rats can be drawn and enable such findings to be extrapolated to human pathology.

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References

- Armario A, Lopez-Calderon A, Jolin T, Castellanos JM. Sensitivity of anterior pituitary hormones to graded levels of psychological stress. Life Sci 1986;39:471-5.
- Black PH, Garbutt LD. Stress, inflammation and cardiovascular disease. J Psychosom Med 2002;52:1–23.
- Blumenthal JA, Jiang W, Waugh RA, Frid DJ, Morris JJ, Coleman E, et al. Mental stress-induced ischemia in the laboratory and ambulatory ischemia during daily life. Association and hemodynamic features. Circulation 1995;92:2102–8.
- De Oliveira AC, D'Almeida V, Hipólide DC, Nóbrega JN, Tufik S. Sleep deprivation decreases total plasma homocysteine levels in rats. Can J Physiol Pharmacol 2002;80:193–7.
- Dicker-Brown A, Fonseca VA, Fink LM, Kern PA. The effect of insulin in the activity of methylenetetrahydrofolate reductase and cystathionine-beta-synthase: studies in hepatocytes. Atherosclerosis 2001;158: 297–301.
- Dierkes J, Jeckel A, Ambrosch A, Westphal S, Luley C, Boeing H. Factors explaining the difference of total homocysteine between men and women in the European Investigation into Cancer and Nutrition Potsdam study. Metabolism 2001;50:640–5.
- Er H, Evereklioglu C, Cumurcu T, Türköz Y, Özerol E, Şahin K, et al. Serum homocysteine level is increased and correlated with endothelin-1 and nitric oxide in Behçet's disease. Br J Ophthalmol 2002;86:653-7.
- Farag NH, Barshop BA, Mills PJ. Effects of estrogen and psychological stress on plasma homocysteine levels. Fertil Steril 2003;79:256–60.
- Fauvel JP. Stress mental et système cardiovasculaire. Ann Cardiol Angeiol (Paris) 2002;51:76-80.
- Finkelstein JD, Martin JJ. Homocysteine. J Biochem Cell Biol 2000; 32:385-9.
- Giorgiades A, Sherwood A, Gullette ECD, Babyak MA, Hinderliter A, Waugh R, et al. Effects of exercise and weight loss on mental stress-

induced cardiovascular responses in individuals with high blood pressure. Hypertension 2000;36:171-6.

- Graham IM, O'Callaghan P. Vitamins, homocysteine and cardiovascular risk. Cardiovasc. Drugs Ther 2002;16:383–9.
- Guilland JC, Favier A, Potier de Courcy G, Galan P, Hercberg S. L'hyperhomocystéinémie: facteur de risque cardiovasculaire ou simple marqueur? 2. Données épidémiologiques. Pathol Biol (Paris) 2003;51: 111–21.
- Hackam DG, Anand SS. Emerging risk factors for atherosclerotic vascular disease. A critical review of the evidence. JAMA 2003;290:932–40.
- House JD, Jacobs RL, Stead LM, Brosnan ME, Brosnan JT. Regulation of homocysteine metabolism. Adv Enzyme Regul 1999;39:69–91.
- Jacobs RL, Stead LM, Brosnan ME, Brosnan JT. Plasma homocysteine is decreased in the hypothyroid rat. Can J Physiol Pharmacol 2000;78: 565–70.
- Jacobs RL, Stead LM, Brosnan ME, Brosnan JT. Hyperglucagonemia in rats results in decreased plasma homocysteine and increased flux through the transulfuration pathway in liver. J Biol Chem 2001;276: 43740–7.
- Lenox RH, Kant GJ, Sessions GR, Pennington LL, Mougey EH, Meyerhoff JL. Specific hormonal and neurochemical responses to different stressors. Neuroendocrinology 1980;30:300–8.
- Meyerhoff JL, Oleshansky MA, Kalogeras KT, Mougey EH, Chrousos GP, Granger LG. Neuroendocrine responses to emotional stress: possible interactions between circulating factors and anterior pituitary hormone release. Adv Exp Med Biol 1990;274:91–111.
- Møller P, Wallin H, Knudsen LE. Oxidative stress associated with exercise, psychological stress and life-style factors. Chem Biol Interact 1996;102: 17–36.
- Muller JR, Le KM, Haines WR, Gan Q, Knuepfer MM. Hemodynamic response pattern predicts susceptibility to stress-induced elevation in arterial pressure in the rat. Am J Physiol 2001;281:R31–7.
- Nygård O, Volsset SE, Refsum H, Brattstrom L, Ueland PM. Total homocysteine and cardiovascular disease. J Intern Med 1999;246:425–54.

- Pacak K, Palkovits M, Yadid G, Kvetnansky R, Kopin IJ, Goldstein DS. Heterogeneous neurochemical responses to different stressors: a test of Selye's doctrine of nonspecificity. Am J Physiol 1998;275:R1247–55.
- Pacák K, Palkovits M. Stressor specificity of central neuroendocrine responses: implications for stress-related disorders. Endocr Rev 2001;22: 502–48.
- Palma BD, Gabriel Jr A, Bignotto M, Tufik S. Sleep deprivation increases endothelin-1 levels. Braz. J Med Biol Res 2002;35:75–9.
- Pfeiffer CM, Huff DL, Gunter EW. Rapid and accurate HPLC assay for plasma total homocysteine and cysteine in a clinical laboratory setting. Clin Chem 1999;45:290–2.
- Ramey SL. Cardiovascular disease risk factors and the perception of general health among male law enforcement officers: encouraging behavioral change. AAOHN J 2003;51:219–26.
- Ratnam S, Maclean KN, Jacobs RL, Brosnan ME, Kraus JP, Brosnan JT. Hormonal regulation of cystathionine beta-synthase expression in liver. J Biol Chem 2002;277:42912–8.
- Stead LM, Brosnan ME, Brosnan JT. Characterization of homocysteine metabolism in the rat liver. Biochem J 2000;350:685–92.
- Stoney CM. Plasma homocysteine levels increase in women during psychological stress. Life Sci 1999;64:2359–65.
- Stoney CM, Engebretson TO. Plasma homocysteine concentrations are positively associated with hostility and anger. Life Sci 2000;66: 2267–75.
- Tobeña R, Horikawa S, Calvo V, Alemany S. Interleukin-2 induces γ-Sadenosyl-L-methionine synthetase gene expression during T-lymphocyte activation. Biochem J 1996;319:929–33.
- Vitvitsky V, Mosharov E, Tritt M, Ataullakhanov F, Banerjee R. Redox regulation of homocysteine-dependent glutathione synthesis. Redox Rep 2003;8:57–63.
- Yudkin JS, Kumari M, Humphries SE, Mohamed-Ali V. Inflammation, obesity, stress and coronary heart disease: is interleukin-6 the link? Atherosclerosis 2000;148:209–14.